

8 HL IB Biology



Gas Exchange

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Gas Exchange in Organisms

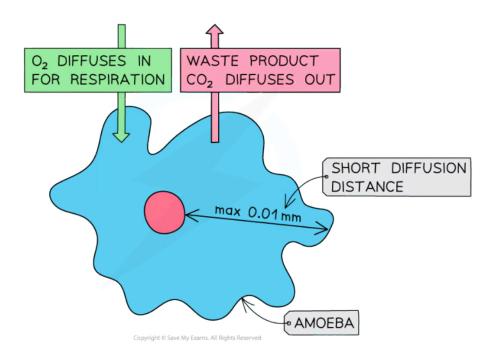
Your notes

Gas Exchange in Organisms

- Cellular respiration is a process occurring in all living cells that releases energy in the form of ATP
 - This energy is released when substrate molecules such as glucose is **oxidised**
 - Organisms use this energy to perform important life functions such as nutrition and excretion
- Aerobic respiration requires oxygen to occur and it produces carbon dioxide as a waste product
 - Living organisms acquire this oxygen from their environment and release carbon dioxide back into their surroundings
- The process by which these gases are exchanged between living organisms and their environment is called gas exchange
 - This includes oxygen uptake and the release of carbon dioxide by organisms
 - In plants, carbon dioxide will be absorbed and oxygen released during the day as a result of photosynthesis
- Gas exchange takes place by the process of diffusion, the rate of which is determined by the following factors:
 - Size of the respiratory surface the bigger the surface, the higher the rate of diffusion
 - Concentration gradient
 - Diffusion distance the shorter the distance, the higher the rate of diffusion
- Small, unicellular organisms such as amoeba have a large surface area compared to the volume of cytoplasm and a short diffusion distance
 - This means that the rate of diffusion is sufficient to supply the organism with enough oxygen to function

Single Celled Organism Diffusion Diagram







Small, unicellular organisms have a large surface area to volume ratio and a short diffusion distance to allow for effective gas exchange to occur

Challenges of gas exchange in organisms

- As an organism increases in size, the challenges of gas exchange become greater
- This is because an **increase in size** will result in a:
 - Smaller surface area to volume ratio
 - Greater diffusion distance
- Large, multicellular organisms therefore cannot rely on diffusion alone to supply every cell with oxygen
 - Another challenge is that the external surface of these organisms are designed to provide protection to the tissue underneath and is therefore not suitable as a respiratory surface
- The cells of **large**, **active** organisms will require **more oxygen** than smaller, less active organisms in order to meet their metabolic demands
 - These organisms will require **specialised organs** for gas exchange

Examiner Tip

Make sure that you do not confuse respiration and gas exchange with each other. Respiration is a chemical process occurring in all living cells while gas exchange refers to the diffusion of oxygen and carbon dioxide across a respiratory surface.



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Gas Exchange Surfaces: Properties

- To **maximise the rate of diffusion** of oxygen and carbon dioxide, gas exchange surfaces require certain properties which include:
 - **Permeability** in order for gases to move across the surface
 - Thin tissue layer to create a short diffusion distance for oxygen and carbon dioxide
 - Presence of **moisture** so that gases can dissolve
 - This will facilitate the diffusion of gases across a gas exchange surface
 - Large surface area so that many gas molecules can diffuse across at the same time





Maintaining a Concentration Gradient

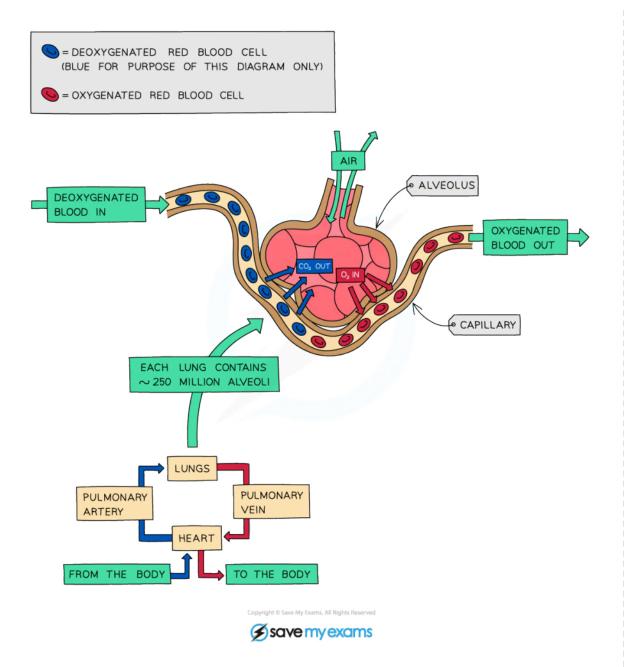
- A steep concentration gradient will ensure a high diffusion rate across a gas exchange surface
 - In organisms, this will allow the diffusion of oxygen into the body and the diffusion of carbon dioxide out of the body
- These concentration gradients are maintained in the following ways:
 - A dense network of blood vessels to provide a large surface area for the diffusion of gases
 - Blood provides a good transport medium for both oxygen and carbon dioxide
 - A continuous blood flow in the blood vessels to ensure that oxygen is constantly transported away from the gas exchange surface and carbon dioxide towards them
 - This ensures that oxygen will always diffuse into the blood and carbon dioxide out of the blood in the lungs
 - **Ventilation** with air in lungs and water in gills to bring oxygen close to the gas exchange surface and to remove carbon dioxide

Alveolus Diagram









The alveolus is the gas exchange surface in humans where a concentration gradient for oxygen and carbon dioxide is maintained



Mammalian Lungs: Adaptations

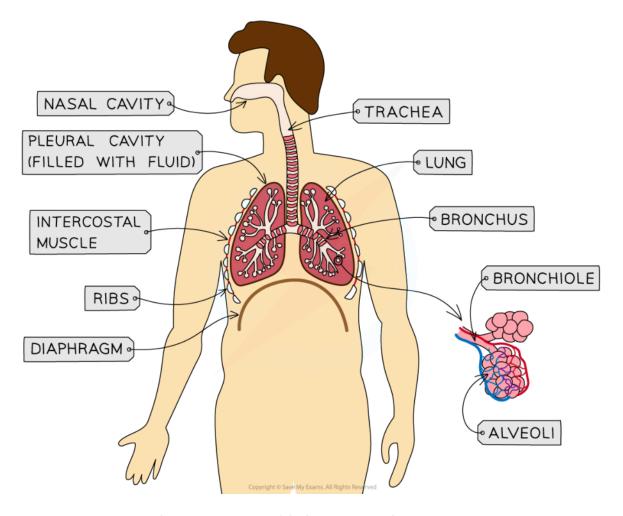
Your notes

Mammalian Lungs: Adaptations

- Air moves in through the nose and mouth before it is carried to the lungs through the trachea
- The **trachea** is a tube supported by **rings of cartilage** which help to support its shape and ensure it stays open while allowing it to move and flex with the body
- The **trachea** divides to form the two **bronchi** (singular bronchus) with walls also strengthened with cartilage and a layer of smooth muscle that can **contract** or **relax** to change the diameter of the airways. Both trachea and bronchi are lined with **ciliated epithelium** to remove particles trapped in mucus that enter the airways
 - One bronchus leads to each lung
- Bronchioles branch off the two bronchi to form a network of narrow tubes
 - The walls of the bronchioles are lined with a layer of smooth muscle to alter the diameter of the bronchiole tubes
 - This helps to regulate the flow of air into the lungs by dilating when more air is needed and constricting when e.g. an allergen is present
- Groups of alveoli are found at the end of the bronchioles
- Each alveolus is surrounded by an extensive network of capillaries to provide a good blood supply for maximum gas exchange

Human Gas Exchange System Diagram





The main structures of the human gas exchange system

Adaptations of mammalian lungs for gas exchange

- Each mammalian lung is comprised of many, small alveoli
 - These provide a large surface area for gas exchange
- Alveoli are grouped around the ends of bronchioles, which spreads out to form a branched network across each lung
 - This ensures an **even distribution** of alveoli throughout the lungs
- The clusters of alveoli are surrounded by an **extensive capillary bed**
 - This provides an **increased surface area** for the diffusion of oxygen and carbon dioxide between the alveoli and blood
 - Deoxygenated blood enters the capillary beds from a branch of the pulmonary artery while oxygenated blood leaves the capillary beds via a branch of the pulmonary vein
 - This maintains the concentration gradient of oxygen and carbon dioxide between the alveoli and blood

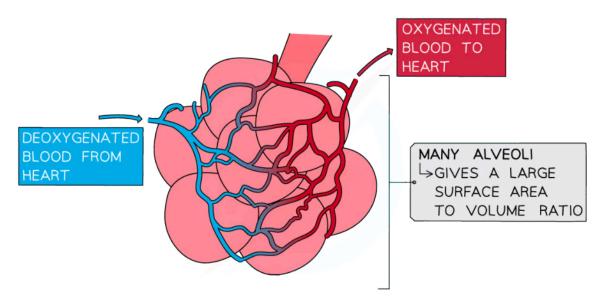


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- Cells of the alveolar wall secrete a substance called **surfactant** which lowers the surface tension in the alveoli
 - This prevents the alveoli from collapsing and sticking together during expiration

Your notes

Human Alveoli Diagram



Many, small alveoli and an extensive capillary network are examples of how the mammalian lung is adapted for gas exchange



Make sure of the terminology that you use here; do not confuse the alveolar wall with a cell wall. The **alveolar wall** is a single layer of epithelial cells that forms the alveoli, while a **cell wall** is a rigid structure that surrounds a plant cell.



Mechanism of Ventilation

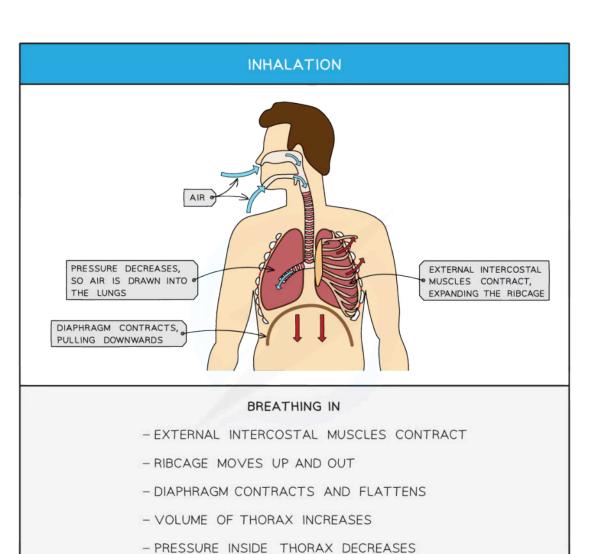
Your notes

Ventilation: Mechanism

- Ventilation is essential for the effective **exchange of gases** in the lungs
 - It replaces older air in the lungs with fresh air from the external environment
 - This helps to maintain the concentration gradient of oxygen and carbon dioxide between the alveoli and blood
- Ventilation involves inspiration (breathing in) and expiration (breathing out)

Inspiration

- The breathing-in, or **inspiration**, process causes the **volume of the chest to increase** and the **air pressure to decrease** until it is **lower than the atmospheric pressure**
 - When gas is in a large volume container that allows the gas particles to spread out, the pressure exerted by the gas on the walls of the container is low
- As a result, air moves **down the pressure gradient** and rushes into the lungs
 - A gas will always move down a pressure gradient from an area of high pressure to an area of low pressure
- The inspiration process
 - The diaphragm **contracts** and **flattens**, increasing chest volume
 - In addition to the flattening of the diaphragm the external intercostal muscles contract, causing the ribcage to move upwards and outwards; this also increases chest volume





The process of inspiration

Expiration

 Breathing out, or expiration, occurs mostly due to the recoil of the lungs after they have been stretched by the inspiration process, and is therefore a mainly passive process

- AIR IS DRAWN IN

 Volume of the chest decreases and pressure increases, causing air to be forced out down its pressure gradient





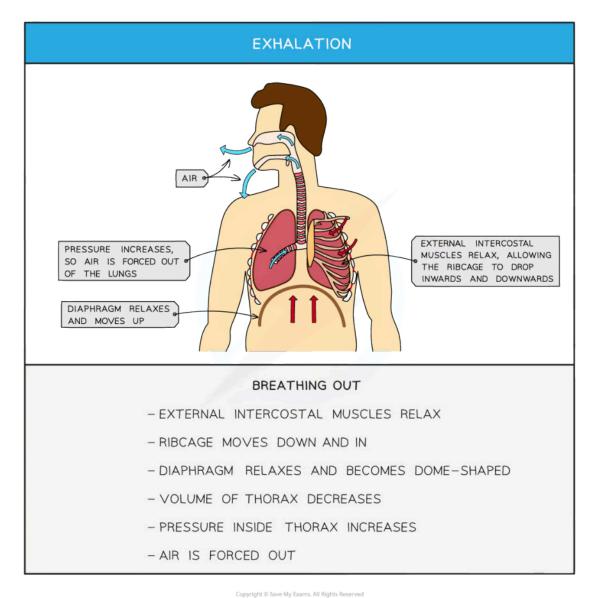
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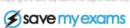
- When gas is in a low volume container it is compressed, causing the gas particles to exert more pressure on the walls of the container
- The passive expiration process
 - External intercostal muscles **relax**, allowing the ribcage to move **down** and **in**
 - Diaphragm relaxes and becomes dome-shaped
 - The **recoil** of **elastic fibres** in the alveoli walls reduces the volume of the lungs
- The expiration process can be active when there is a need to expel excess air from the lungs e.g. when blowing out a candle
- The active expiration process
 - Internal intercostal muscles contract to pull the ribs down and in
 - Abdominal muscles contract to push organs upwards against the diaphragm, decreasing the volume of the chest cavity
 - This causes forced exhalation











The process of passive expiration



Measuring Lung Volumes: Skills

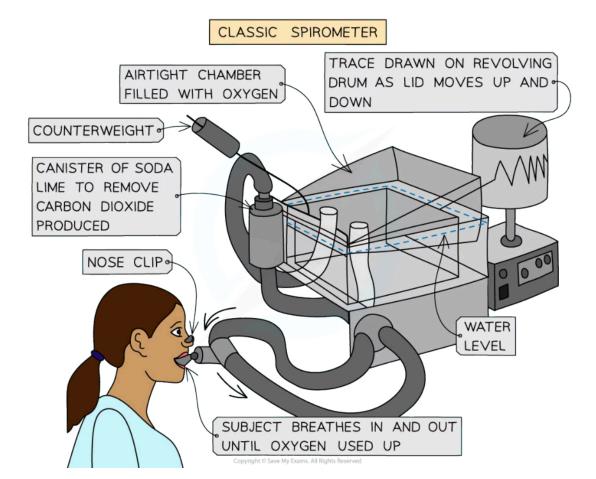
Your notes

Measuring Lung Volumes

- It is possible to investigate the effect of exercise on ventilation using an apparatus called a **spirometer**
 - It contains a chamber filled with water which is covered by a hinged plastic lid
 - The person partaking in the experiment breathes through a mouthpiece which is connected to the spirometer chamber
 - The plastic lid moves up and down as breathing occurs
- The spirometer chamber could be filled with either air or oxygen
 - When filled with air, it can be used to determine lung capacity in different conditions
 - When filled with oxygen and soda lime (for absorbing carbon dioxide), it can measure oxygen consumption in different conditions
- Spirometer traces are created by:
 - Drawing a line on a revolving drum as the lid moves
 - A computer which draws a graph of the results
- Several **measurements can be made** using spirometer traces such as:
 - Ventilation rate
 - Tidal volume
 - Reserve volumes during inspiration and expiration
 - Vital capacity



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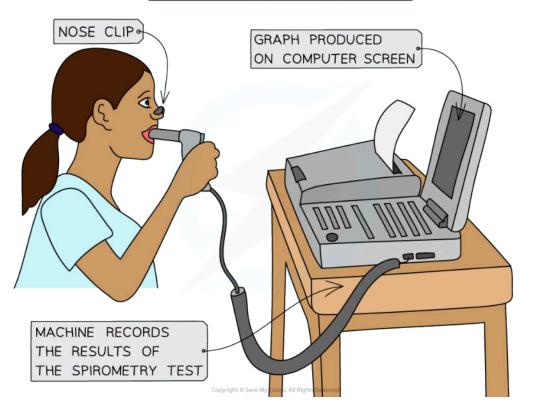


A classic spirometer can be used to investigate ventilation



FULLY ELECTRONIC SPIROMETER

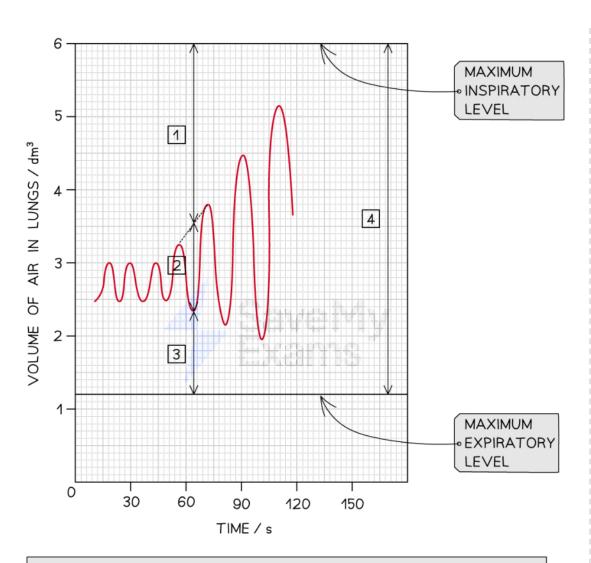




Using a spirometer to monitor ventilation can also be carried out with an electric spirometer

Analysis of spirometer trace

• The effect of exercise on ventilation can be seen in the spirometer trace below



Your notes

KEY: 1 = INSPIRATORY LEVEL 3 = EXPIRATORY RESERVE VOLUME 2 = TIDAL VOLUME 4 = VITAL CAPACITY

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Tidal volume

- The **tidal volume** is the volume of air inhaled and exhaled during normal breathing
 - Exercise will lead to an increase in the tidal volume as more air is moved in and out of the lungs
 - We do have the potential to take extra deep breaths
 - The maximum volume of air that can enter the lungs during inspiration is known as the maximum inspiratory level
 - Similarly, the maximum volume of air that can be exhaled during expiration is known as the maximum expiratory level

Inspiratory and expiratory reserve volumes



- The **reserve volumes** of the lungs refer to the extra volume of air that can be inhaled or exhaled when taking an extra deep breath and are determined as follows:
 - The difference between the maximum inspiratory level and tidal volume is called the **inspiratory** reserve volume
 - The difference between the maximum expiratory level and tidal volume is called the expiratory reserve volume

Vital capacity

- The vital capacity (VC) refers to the total amount of air exhaled after taking a deep breath
 - This can be calculated by adding the **tidal volume** (**TV**), **inspiratory reserve volume** (**IRV**) and **expiratory reserve volume** (**ERV**) together

VC = TV + IRV + ERV

Ventilation rate

- The ventilation rate can be determined by counting the number of inhalations or exhalations per minute
 - Exercise will cause an increase in the ventilation rate as you will be taking more breaths per minute





Gas Exchange in Plants

Your notes

Leaf Adaptations for Gas Exchange

- Gas exchange in plants occur through the leaf
- The leaf contains the following tissues:
 - Epidermal tissue forming the outer boundary of the leaf
 - Mesophyll tissue that make up the bulk of internal structure of the leaf
 - Vascular tissue which transports substances between the leaf and the rest of the plant

Epidermis

- This is formed by a single layer of tightly packed cells
 - The leaf has an upper and lower epidermis which protects the inner parts of the leaf
- The lower epidermis contains tiny pores called **stomata** (singular stoma)
 - Each stoma is surrounded by two guard cells which controls the opening and closure of the pore
 - When water moves into the guard cells they become turgid and change shape which opens the stomata
 - They become flaccid when water is lost and this causes the stomata to close
 - Stomata are the structures through which gas exchange occur in a leaf
 - They allow for the diffusion of oxygen and carbon dioxide into and out of the leaf
- The epidermis is often covered by a waxy layer called the **cuticle**
 - This forms an impermeable barrier

Mesophyll tissue

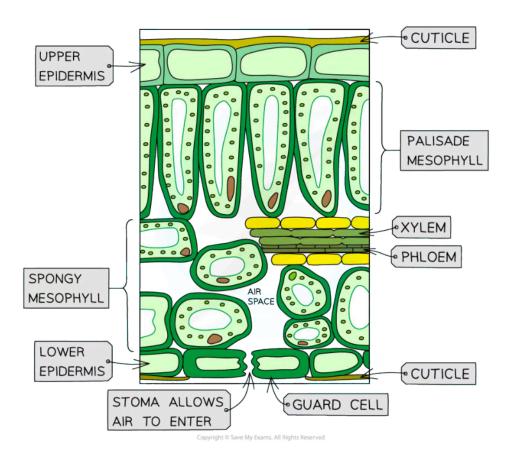
- These are formed by **parenchyma cells** which contain **chloroplasts**
 - This is where photosynthesis occurs in the leaf
- Two types of mesophyll tissue are found in the leaf:
 - Palisade mesophyll forms a layer beneath the upper epidermis and contain many chloroplasts for maximum photosynthesis
 - Spongy mesophyll contains large air spaces between the cells for gas exchange to occur

Vascular tissue

- Vascular tissue is arranged in vascular bundles and is responsible for the transport of substances around the plant
 - Vascular bundles form the veins in leaves
 - **Xylem** transports **water** and mineral ions from the roots to the leaves
 - Phloem transports the products of photosynthesis from the leaves to other parts of the plant

Structure of a Leaf Diagram







The structure of a leaf has distinct layers each with their own function

Adaptations for gas exchange

• The leaf has several adaptations that facilitate gas exchange

Leaf Adaptations for Gas Exchange Table

Adaptation	Function
Waxy cuticle	Prevents gases and water vapour from leaving through the epidermis so that gas exchange must occur through stomata. This allows gas exchange and water loss to be controlled
Epidermis	Contain stomata for gas exchange. Most stomata are found in the lower epidermis where the temperature is lower. This reduces water loss
Air spaces	Maintains a concentration gradient of gases between the air and spongy mesophyll cells by allowing movement of gases
Spongy mesophyll	Increases the surface area for gas exchange

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Guard cells	Control gas exchange and water loss by opening or closing stomata
Veins	Xylem vessels bring water to the leaf which is required for photosynthesis and transpiration. Photosynthesis requires carbon dioxide to diffuse into the leaf while transpiration involves the loss of water vapour

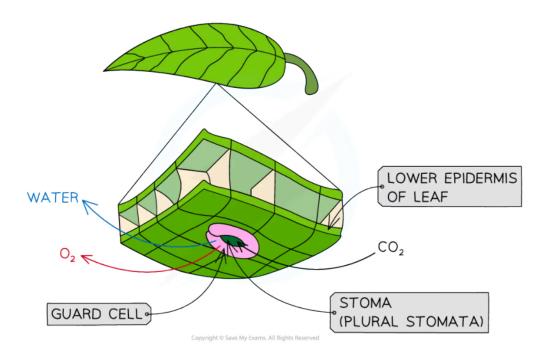




Transpiration: Consequence of Gas Exchange

- The majority of photosynthesis takes place in the **leaves** of plants
 - Some plants are able to carry out photosynthesis in the cells of their stems
- During photosynthesis, carbon dioxide is taken in by the leaf and oxygen is released
 - The **pores in the epidermis of the leaf** through which this gas exchange takes place are known as stomata (singular stoma)
 - The stomata need to be open all the time in order for gas exchange, and therefore photosynthesis, to continue
- The problem for plants is that as the stomata open to allow gas exchange to occur, water in the form of water vapour is also lost through the stomata
 - This water loss is known as **transpiration**
 - Most plants can use cells called guard cells to close their stomata in order to reduce water loss,
 but this will also reduce gas exchange and therefore their rate of photosynthesis
 - Transpiration is the inevitable consequence of gas exchange in the leaf
- There are some **advantages** to the process of transpiration
 - It provides a means of cooling the plant via evaporation
 - The transpiration stream is helpful in the uptake of mineral ions
 - The turgor pressure of the cells, due to the presence of water as it moves up the plant, provides
 support to the leaves and to the stems of non-woody plants
 - Leaves with high turgor pressure do not wilt and therefore have an increased surface area for photosynthesis

Transpiration in the Leaf Diagram







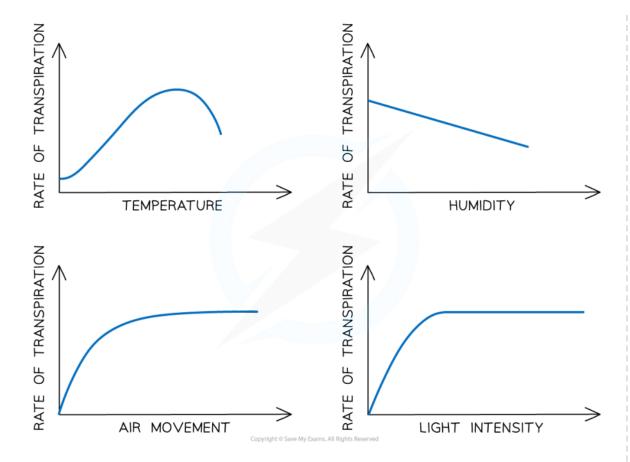
The loss of water vapour from leaves by evaporation through the stomata is unavoidable as gas exchange for photosynthesis can only occur when the stomata are open

Your notes

Factors affecting the rate of transpiration

- Air movement
 - More air movement leads to increased rates of transpiration
 - The air outside a leaf usually contains a lower concentration of water vapour than the air spaces inside a leaf, causing water vapour to diffuse out of the leaf
 - When the air is relatively still, water molecules can accumulate just outside the stomata, creating a local area of high humidity
 - Less water vapour will diffuse out into the air due to the reduced concentration gradient
 - Air currents, or wind, can carry water molecules away from the leaf surface, increasing the concentration gradient and causing more water vapour to diffuse out
- Temperature
 - Higher temperatures lead to higher rates of transpiration, up to a point at which transpiration rates will slow
 - An increase in temperature results in an increase in the kinetic energy of molecules
 - This increases the rate of transpiration as water molecules evaporate out of the leaf at a faster rate
 - If the temperature gets too high the stomata close to prevent excess water loss
 - This dramatically reduces the rate of transpiration
- Light intensity
 - Higher light intensities will increase the rate of transpiration up to a point at which transpiration
 rates will level off
 - Stomata close in the dark and their closure greatly reduces the rate of transpiration
 - **Stomata open when it is light** to enable gas exchange for photosynthesis; this increases the rate of transpiration
 - Once the stomata are all open any increase in light intensity has no effect on the rate of transpiration
- Humidity
 - Higher humidity levels reduce the rate of transpiration
 - If the humidity is high that means the air surrounding the leaf surface is saturated with water vapour
 - This causes the rate of transpiration to decrease as there is no concentration gradient between the inside of the leaf and the outside
 - At a certain level of humidity, an **equilibrium** is reached; water vapour levels inside and outside the leaf are the same, so there is no net loss of water vapour from the leaves







Several environmental factors affect the rate of transpiration in plants

Examiner Tip

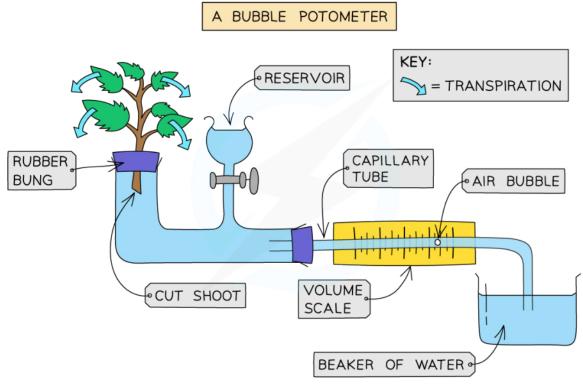
Take note that the movement of water molecules during transpiration is **not** by osmosis. One of the requirements of osmosis is that water molecules move across a cell membrane, which does not happen during transpiration. We therefore say that water vapour **diffuses** out of the leaf through stomata during transpiration

Measuring the rate of transpiration

- The **effect of environmental factors on the rate of transpiration** in plants can be measured using a piece of equipment called a **potometer**
 - Note that while potometers are used to measure transpiration rates, they technically measure the
 rate of water uptake rather than the rate of transpiration, as a small amount of the water taken up
 by a plant will be used in photosynthesis
 - Because the amount of water used in photosynthesis is so small in relation to the total amount
 of water that passes through a plant, the rate of water uptake can reasonably be used to
 represent the rate of transpiration



- Different types of potometer exist
 - Bubble potometers measure the movement of an air bubble along a water-filled tube connected to a plant shoot as water is drawn up by the shoot
 - The position of the air bubble is recorded at the start of an experiment, and then a researcher can either measure how far the bubble moves in a set amount of time, or time how long it takes for the bubble to move a certain distance
 - Mass potometers measure the change in mass of a water-filled test tube connected to a plant shoot as it loses water over a set amount of time
- The effect of **various environmental factors** on transpiration can be measured by placing the potometer in different conditions e.g.
 - Wind speed
 - Humidity
 - Light intensity
 - Temperature



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A bubble potometer uses the movement of an air bubble to measure the rate at which water is drawn up by a plant shoot. In this image the air bubble will move to the left along the tube as the plant transpires

- Environmental factors can be investigated in the following ways
 - Air movement
 - A fan on different settings could be used to vary the flow of air around a plant shoot





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- Humidity
 - Enclosing the plant shoot in a **plastic bag** can increase the humidity
 - A humidifier or dehumidifier could be used to give a measurable variation in humidiy
- Light intensity
 - A lamp at different distances or with different types of light bulb can be used to vary light intensity
- Temperature
 - A thermometer or temperature probe can be used to find surroundings with different air temperatures
 - A heater or air conditioner can be used to give a measurable variation in temperature
- A researcher would need to be aware of the importance of controlling any variables other than the variable being investigated to ensure that any results are valid e.g. placing a plant shoot in different rooms could be a way of varying temperature, but might bring the risk of also varying light levels and humidity; these variables would need to be controlled





Drawing Leaf Structure: Skills

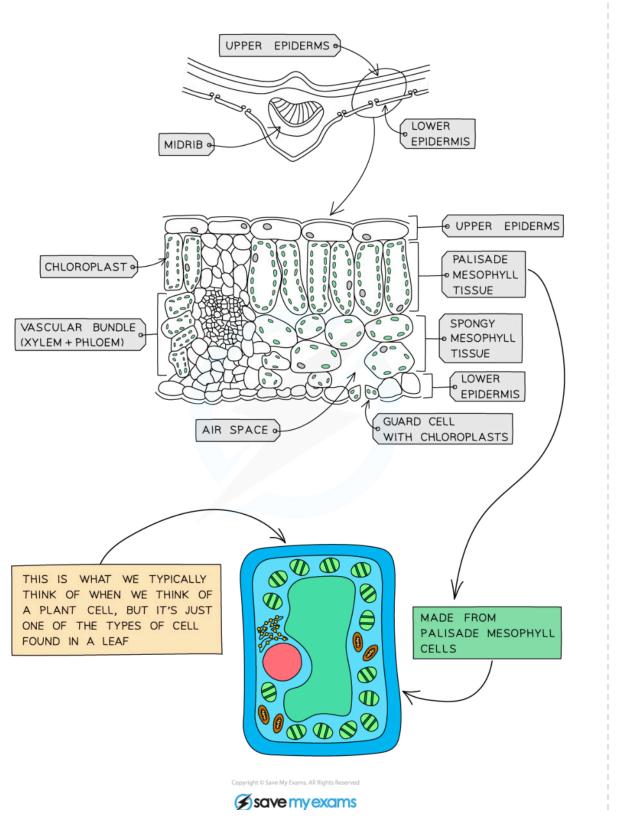
Your notes

Drawing Leaf Structure

- You will be expected to identify the following structures in the leaf of a dicotyledonous plant:
 - Chloroplasts
 - Cuticle
 - Guard cells
 - Stomata
 - Upper and lower epidermis
 - Palisade mesophyll
 - Spongy mesophyll
 - Air spaces
 - Vascular bundles (xylem and phloem)

Structure of Leaf Diagram







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Diagram showing the transverse section of a leaf

Your notes

Drawing a plan diagram

- Plan diagrams are drawings made from micrographs or from viewing specimens under a low magnification
- Keep the following in mind when drawing a plan diagram:
 - No individual cells are drawn, only tissue layers enclosed by lines should be present
 - Pay attention to the **distribution of tissue** throughout the plant organ
 - Use a sharp pencil and draw clear, continuous lines
 - Do not shade any part of your drawing
 - Make sure your **proportions and observations** are accurate
 - Draw what you actually see, not what you would expect to see from a textbook
 - Draw your drawing **big enough** to fill up at least half the available space
- When labelling your plan diagram remember to:
 - Use a ruler to draw label lines, not freehand
 - Avoid using arrowheads and make sure the label lines stop at the structure
 - Make sure label lines **do not cross** each other
 - Write all labels **horizontally**, not at different angles





Worked example

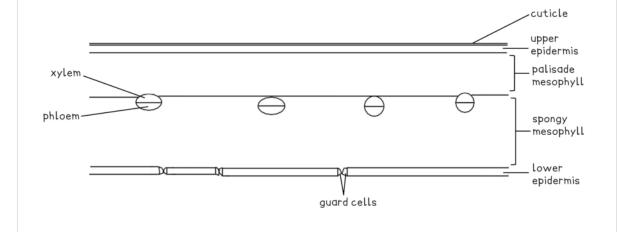
The following micrograph shows a transverse section of a dicotyledonous leaf.

Draw a labelled plan diagram of this micrograph.



By Berkshire Community College Bioscience Image Library, Public domain, Wikimedia

Answer:







Determining Stomatal Density: Skills

Your notes

Determining Stomatal Density

- The **density of stomata** (the number of stomata per unit of area) can be a useful measurement to biologists
 - To assess the plant's likely response to a dry spell of weather
 - To predict its behaviour in windy or wet climates if the plant was being moved for agricultural / horticultural reasons
- This technique can be used to assess how stomatal density varies from species to species

Apparatus

- A plant to sample a leaf from
- Clear nail varnish (ideally solvent based)
- Sellotape
- Microscope
- Microscope slides
- Stage micrometer
- Counting device (clicker/ phone app etc.)
- Calculator

Method

- Select a leaf from a live plant and cut it off the plant
 - Geraniums and spider plants make good subjects for this experiment
- Place the leaf upside down on a flat surface such as a tile or worktop
- Paint clear nail varnish onto the underside of the leaf
- Wait for the nail varnish to **dry** (approx. 5 minutes)
- **Peel off** the layer of varnish using sellotape
 - Discard the leaf
 - The layer of varnish now forms a **leaf cast**
- Place the dried varnish impression on a **microscope slide**
 - A coverslip is not required as this isn't a biological sample, just an impression of one
 - A drop of water is not required either, so long as the sample is laid flat on the slide
- Use the usual steps to focus on the sample (you can read about this in our revision note on microscope skills)
- Adjust the zoom such that a countable number of stomata are visible in the field of view
 - Between **15 and 100** is ideal
 - Even if a stoma is partially visible at the edge, still count it as 1
- Count the stomata in that field of view
 - You may wish to use a **clicker** or **phone app** so you don't lose count!
- Move the field of view to another area of the nail varnish layer and repeat
- Count at least 3 separate fields of view and take a mean value
 - Repeat readings allow you to eliminate anomalous results and calculate a reliable mean



Measurements to take

- Use a **stage micrometer** to measure the **diameter** of the field of view
 - This has to be at the same magnification power that you used when counting the stomata
- The stage micrometer will be calibrated in micrometers
 - A typical microscope allows the scientist to look at a field of view of about 0.5 mm diameter when on full power (x 400)
- You will have calculated a mean number of stomata per field of view from the previous stage
- You can read about using a stage micrometer in our revision notes on microscope skills





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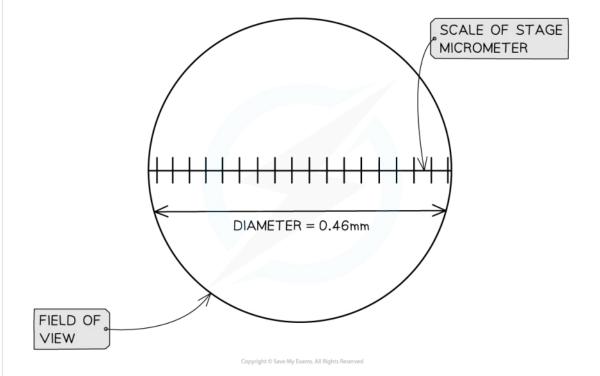


Worked example

A study reveals a mean count of 16 stomata per field of view at a magnification of \times 400. The stage micrometer calculates the **diameter** of the field of view at a magnification of \times 400 to be **0.46mm**

Calculate the stomatal density based on these data. Give units in stomata per mm^2

Use a value of π = 3.14 and give your answer to the nearest whole number of stomata.



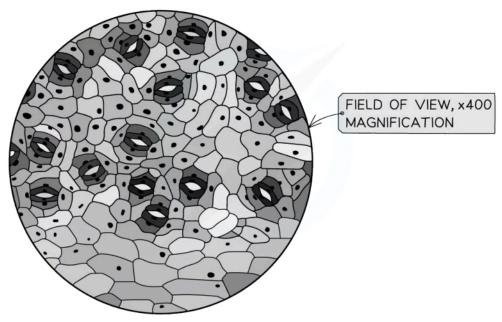




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COUNT = 16 STOMATA IN THIS FIELD OF VIEW





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Answer:

Step 1: Calculate the radius of the field of view

Radius = Diameter \div 2

Radius = $0.46 \text{ mm} \div 2 = 0.23 \text{ mm}$

Step 2: Calculate the area of the field of view

Area = $\pi r^2 = \pi \times 0.23^2$

Area = $0.1662 \, \text{mm}^2$

Step 3: Divide the mean number of stomata by the area of the field of view to calculate density

Density = $16 \div 0.1662 = 96.27$ stomata per mm²

Step 4: Round to the required precision (nearest whole number)

Density = 96 stomata per mm²

Limitations

• Not all plant species have easily accessible stomata that create a strong imprint



- Solvent-based nail varnish can destroy some of the cell structure it comes into contact with
- Does the plant grow more stomata (guard cells) according to the conditions in each individual habitat?
- Water-based nail varnish is safer to use but dries **more slowly**

Your notes

NOS: Reliability of quantitative data is increased by repeating measurements

- Reliability refers to the level of trust that we can place in numerical measurements
 - These types of measurements are known as quantitative data
- Repeating the stomatal count for the same species of leaf under the same conditions will increase the reliability of the results
 - It is very possible that the data gathered during a single count could contain errors that we may not be aware of
 - Taking repeated measurements will identify anomalous measurements and allow us to calculate a mean
 - Anomalous measurements are those that **deviate** from the expected measurements
 - Anomalies are **omitted** when calculating the mean for a data set
- If repeated stomatal counts deliver similar results, the data is said to be **reliable**
 - We can therefore place a higher level of trust in the data than what would have been possible if we got very different results with every count
- Repeating measurements is a crucial step in gathering data during a scientific investigation



Anomalous results are sometimes referred to a outliers.



Haemoglobin & Oxygen (HL)

Your notes

Foetal & Adult Haemoglobin

- Haemoglobin is the molecule responsible for **binding oxygen** in our blood
 - They are **globular proteins** found in abundance in red blood cells
 - Each haemoglobin molecule consists of four polypeptide subunits
 - At the centre of each subunit is an **iron-containing haem group** with which oxygen combines
 - Each haem group can bind to **one** oxygen molecule
 - That means that each molecule of haemoglobin can transport four oxygen molecules
- Oxygen is one of the gases found in air and each of these gases exerts a pressure
 - The pressure of each gas in a mixture of gases is called its **partial pressure**
 - The symbol for partial pressure is p, therefore the partial pressure of oxygen can be denoted as pO_2
- Due to the shape of the haemoglobin molecule it is difficult for the first oxygen molecule to bind to its haem group
- However, after the first oxygen molecule binds, the haemoglobin protein changes shape, or conformation, making it easier for the next oxygen molecules to bind
 - This is known as **cooperative binding**
- The ease with which haemoglobin binds and dissociates with oxygen can be described as its affinity for oxygen
 - In areas where there are **high partial pressures** of oxygen (such as the alveoli of the lungs), the affinity of haemoglobin for oxygen is **high**
 - This means haemoglobin and oxygen will **bind easily**
 - In areas where there are low partial pressures of oxygen (such as respiring muscle cells), the affinity of haemoglobin for oxygen is low
 - This means haemoglobin and oxygen will dissociate easily from each other
 - This ensures that haemoglobin can easily bind to oxygen in the lung capillaries to transport it to and then release it near respiring cells that require oxygen

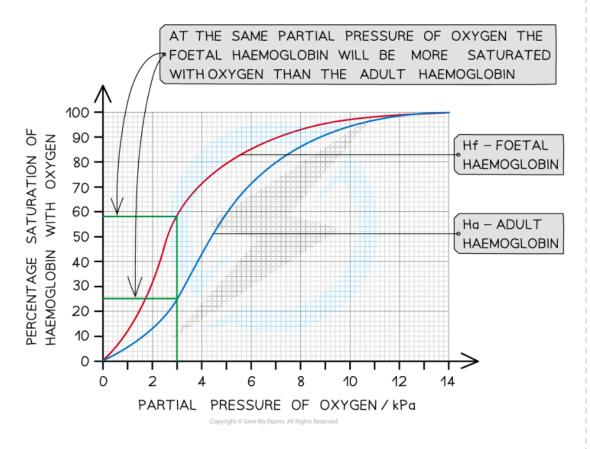
Foetal haemoglobin

- The haemoglobin of a developing foetus has a higher affinity for oxygen than adult haemoglobin
- This is vital as it allows a foetus to obtain oxygen from its mother's blood at the placenta
 - Foetal haemoglobin can bind to oxygen at low pO₂
 - At this low pO₂ the mother's haemoglobin is **dissociating with oxygen**
- We can represent the percentage saturation of haemoglobin at different partial pressures of oxygen as a graph
 - This is called the oxygen dissociation curve
- On a dissociation curve, the curve for foetal haemoglobin shifts to the left of that for adult haemoglobin
 - This means that at any given partial pressure of oxygen, foetal haemoglobin has a higher percentage saturation than adult haemoglobin
- After birth, a baby begins to produce adult haemoglobin which **gradually replaces** foetal haemoglobin



 This is important for the easy release of oxygen in the respiring tissues of a more metabolically active individual





Foetal haemoglobin has a higher affinity for oxygen; its oxygen dissociation curve therefore lies further to the left than the curve of adult haemoglobin

- Haemoglobin has the ability to change shape, or conformation, once oxygen binds to it due to cooperative binding
 - Proteins like this are known as **allosteric proteins** as they can exist in multiple conformations
- Carbon dioxide is an **allosteric inhibitor** of haemoglobin
 - This means that when it binds to haemoglobin, it is more difficult for oxygen to bind to haemoglobin as the protein cannot change its conformation
 - This **lowers the affinity** of haemoglobin for oxygen
- Carbon dioxide has **less** of an allosteric effect on foetal haemoglobin
 - This enables foetal haemoglobin to have a higher affinity for oxygen even if carbon dioxide is bound to it



The Bohr Shift (HL)

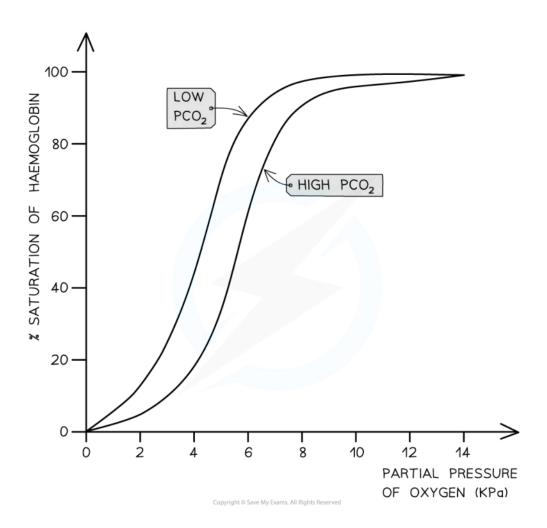
Your notes

The Bohr Shift

- Changes in the oxygen dissociation curve as a result of carbon dioxide levels are known as the Bohr effect, or Bohr shift
- When the partial pressure of carbon dioxide in the blood is high, haemoglobin's affinity for oxygen is reduced
 - This is the case in respiring tissues, where cells are producing carbon dioxide as a waste product
 of respiration
 - This occurs because CO₂ lowers the pH of the blood
 - CO₂ combines with water to form carbonic acid
 - Carbonic acid dissociates into hydrogen carbonate ions and hydrogen ions
 - Hydrogen ions bind to haemoglobin, causing the release of oxygen
- This is a helpful change because it means that **haemoglobin gives up its oxygen more readily** in the respiring tissues where it is needed
- On a graph showing the dissociation curve, the curve **shifts to the right** when CO₂ levels increase
 - This means that at any given partial pressure of oxygen, the percentage saturation of haemoglobin is lower at higher levels of CO₂







The dissociation curve shifts to the right as a result of the Bohr effect. This means that any given partial pressure of oxygen, the percentage saturation of haemoglobin is lower at higher CO_2 levels.

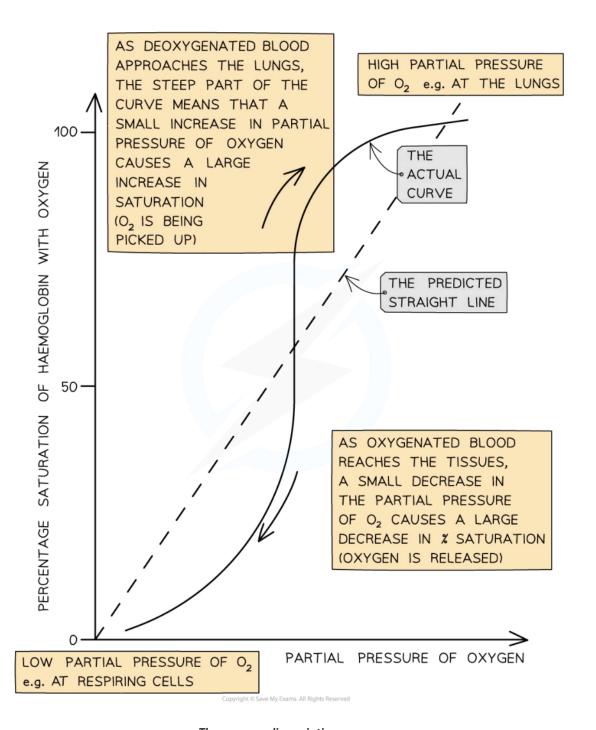


The Oxygen Dissociation Curve (HL)

Your notes

The Oxygen Dissociation Curve

- The oxygen dissociation curve shows the rate at which oxygen associates, and also dissociates, with haemoglobin at different partial pressures of oxygen (pO₂)
 - Partial pressure of oxygen refers to the pressure exerted by oxygen within a mixture of gases; it is
 a measure of oxygen concentration
 - Haemoglobin is referred to as being saturated when all of its oxygen binding sites are taken up with oxygen; so when it contains four oxygen molecules
- The ease with which haemoglobin binds and dissociates with oxygen can be described as its affinity for oxygen
 - When haemoglobin has a high affinity it **binds easily** and **dissociates slowly**
 - When haemoglobin has a low affinity for oxygen it **binds slowly** and **dissociates easily**
- In other liquids, such as water, we would expect oxygen to becomes associated with water, or to dissolve, at a **constant rate**, providing a **straight line** on a graph, but with haemoglobin **oxygen binds at different rates** as the pO₂ changes; hence the resulting curve
 - It can be said that haemoglobin's affinity for oxygen changes at different partial pressures of oxygen



The oxygen dissociation curve

Interpreting the curve





- When the curve is read from left to right, it provides information about the rate at which haemoglobin binds to oxygen at different partial pressures of oxygen
 - At low pO₂ (the bottom left corner of the graph) oxygen binds slowly to haemoglobin; this means
 that haemoglobin cannot pick up oxygen and become saturated as blood passes through the
 body's oxygen-depleted tissues
 - Haemoglobin has a **low affinity for oxygen at low pO**₂, so saturation percentage is low
 - At medium pO₂ (in the central region of the graph) oxygen binds more easily to
 haemoglobin and saturation increases quickly; at this point on the graph a small increase in
 pO₂ causes a large increase in haemoglobin saturation
 - At high pO_2 (in the top right corner of the graph) **oxygen binds easily to haemoglobin**; this means that haemoglobin can pick up oxygen and become saturated as blood passes through the lungs
 - Haemoglobin has a high affinity for oxygen at high pO₂, so saturation percentage is high
 - Note that at this point on the graph increasing the pO₂ by a large amount only has a small
 effect on the percentage saturation of haemoglobin; this is because most oxygen binding
 sites on haemoglobin are already occupied
- When read from right to left, the curve provides information about the rate at which haemoglobin dissociates with oxygen at different partial pressures of oxygen
 - In the lungs, where **pO₂ is high**, there is **very little dissociation** of oxygen from haemoglobin
 - At medium pO₂, oxygen dissociates readily from haemoglobin, as shown by the steep region of the curve; this region corresponds with the partial pressures of oxygen present in the respiring tissues of the body, so ready release of oxygen is important for cellular respiration
 - At this point on the graph a small decrease in pO₂ causes a large decrease in percentage saturation of haemoglobin, leading to easy release of plenty of oxygen to the cells
 - At low pO₂ dissociation slows again; there are few oxygen molecules left on the binding sites, and the release of the final oxygen molecule becomes more difficult, in a similar way to the slow binding of the first oxygen molecule

Explaining the curve

- The curved shape of the oxygen dissociation curve for haemoglobin can be explained as follows
 - Due to the shape of the haemoglobin molecule it is difficult for the first oxygen molecule to bind to haemoglobin
 - This means that binding of the first oxygen occurs slowly, explaining the relatively shallow curve at the bottom left corner of the graph
 - After the first oxygen molecule binds to haemoglobin, the haemoglobin protein changes shape, or conformation, making it easier for the next oxygen molecules to bind due to cooperative binding
 - This speeds up binding of the remaining oxygen molecules and explains the steeper part of the curve in the middle of the graph
 - As the haemoglobin molecule approaches saturation it takes longer for the fourth oxygen molecule to bind
 - This is due to the shortage of remaining binding sites, explaining the levelling off of the curve in the top right corner of the graph

